

# **Algerian Journal of Biosciences**

Journal homepage: http://www.ajbjournal.periodikos.com.br



# A review

# Biological control by Plant Growth Promoting Rhizobacteria

Bestami Merdia a, Ben Malek Rokaia a, Fellan Kheira a and Benaissa Asmaa a.b \*

<sup>a</sup> Department of Biology, Institute of Sciences and Technology, Universitary Center of Amine Elokkal El Hadj Moussa Eg. Akhamoukh, 11039 Sersouf, Tamanrasset, Algeria

b Laboratory of Plant Physiology, Department of Biology and Physiology of Organisms, Faculty of Biological Sciences, University of Sciences and Technologies of Houari Boumediene - El-Alia BP 16011 Bab Ezzouar, Algiers, Algeria

### ARTICLE INFO

### Article history: Received 22 October 2020 Revised 10 December 2020 Accepted 13 December 2020

Keywords: Plant Growth Promoting Rhizobacteria; Biocontrol: Phytopathogen.

### **ABSTRACT**

Plant Growth Promoter Rhizobacteria (PGPR) is soil bacteria that can live on, in or around plant tissue and promote plant growth by many mechanisms that include a biological control of plant pathogens. Indeed, PGPRs have a protective effect through several modes of action such as antagonism, competition, production of hydrolytic enzymes and biofilm formation. Moreover, the use of PGPRs as biocontrol agents is very harmonious with the environment and therefore represents a good alternative to the use of chemicals in agriculture. This review is presented as a general bibliographical synthesis on the different aspects of PGPRs and their biocontrol potential.

© 2020 Faculty of Natural Sciences and Life, University of Echahid Hamma Lakhdar. All rights reserved

### Introduction

Throughout the world, concern about plant diseases that can affect agriculture is becoming more and more serious due to severe crop damage [1]. Post-harvest diseases are a major cause of deterioration of unprocessed fruits and vegetables [47]. Economic losses are enormous [4]. In order to protect crops against plant pathogens and to meet the social and economic needs of the inhabitants, farmers have become dependent on chemicals as a method of crop protection [25]. These chemicals, such as pesticides, are an effective tool to solve these problems, but they have harmful consequences by rapid resistance of plant pathogens to antibiotics [58]. On the other hand, this leads to the creation of multi-resistant germs that spread in microbial populations, which generally attack agriculture and infect humans and animals when they consume these crops, in addition to several other damages to the environment and the ecological balance [58].

In view of the drawbacks of chemical remediation, the scientific community has turned to new control methods less harmful to the environment in order to ensure the sustainability of agriculture by increasing its profitability and safeguarding natural resources for future generations [24], and this has led to the use of antagonistic biological agents found in large numbers in the soil. In fact, the rhizosphere (the part of the soil that surrounds the roots) is an important ecological niche of microbial biodiversity, rich in nutrients due to plant root exudates, and which interacts between it and the roots of host plants [7]. Among these rhizospheric antagonistic agents, PGPRs (plant growth promoting rhizobacteria) are in the front line of attack. They have the capacity to effectively colonize host plant roots and whose overall effect is to promote their directly through soil biofertilization phytostimulation through the production of phytohormones or indirectly through the improvement of stress tolerance and the control of phytopathogens, this phenomenon is commonly referred to as "biocontrol" [57].

The use of PGPR as a biocontrol agent is therefore a hot topic and has been the focus of recent work, as its use is considered the best alternative to chemical control [14]. Thus, biocontrol is based on the use of natural enemies to eliminate aggressors or phytopathogenic agents through

Peer review under responsibility of University of Echahid Hamma Lakhdar. © 2020 University of Echahid Hamma Lakhdar. All rights reserved.

DOI: http://dx.doi.org/10.5281/zenodo.4393567

<sup>\*</sup> Corresponding author: Asmaa BENAISSA. Tel.: 00213556786196 E-mail address: benaissa.asmaa@yahoo.fr

several mechanisms such as antagonism, competition, biofilm formation, as well as the synthesis of hydrolytic enzymes [39].

In this perspective, our work focused on the main mechanisms of action attributed to PGPRs and their roles in the biocontrol of plant diseases.

### 2. General information

### 2.1. Rhizosphere

The bacteriologist and agronomist Lorenz Hiltner were first defined the term rhizosphere in 1904 [30] as a thin layer of soil where interactions between absorbent roots and microorganisms take place [9], with a composition changed by the metabolism and activity of the latter [44].

#### 2.2. Rhizobacteria

According to Kloepper [33], rhizobacteria or rhizospheric bacteria are a specific community of soil bacteria that have the ability to colonize the rhizospheric soil, with the potential to reside in contact with plant roots at various stages of development and growth. Bacteria meeting this definition belong to different genera and species, of which the most studied are *Pseudomonas*, *Azospirillum*, *Agrobacterium* and *Bacillus* [38].

### 2.3. Plant Growth Promoting Rhizobacteria (PGPR)

Plant Growth Promoting Rhizobacteria (PGPR) is a rhizobacteria that has *the* ability to enhance plant growth directly or indirectly through the colonization of root systems [35]. This term was first used by Kloepper and Schroth in 1978 [34] to designate *Pseudomonas fluorescens* strains. Indeed, PGPRs include those that live freely, and those that form specific symbiotic relationships with plants [6].

## 2.4. Mechanisms of action of the PGPRs

PGPRs promote plant growth through several mechanisms, either directly or indirectly (Fig. 1).

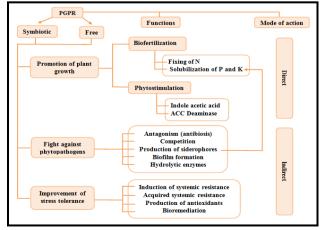


Fig 1. Schematic diagram representing the forms, functions and modes of action of PGPRs [5].

### 2.4.1. Biofertilization

The aim of biofertilizers is the improvement of plant nutrition through biological processes, two of the most relevant of which are:

- (1) The biological fixation of atmospheric nitrogen by reducing it to ammonia through a complex enzyme system mainly nitrogenase [32].
- (2) The phosphate-solubilizing microorganisms (PSM), which are characterized by their ability to solubilize precipitated forms of phosphorus and include a wide range of symbiotic and non-symbiotic organisms [40].

### 2.4.2. Phytostimulation

Phytostimulation is a hormonal stimulation of the plant through the availability of the latter or their precursor by PGPR. We will quote as examples: (1) the production of Acetic Indole Acid (AIA) by some PGPR of the genus Pseudomonas, Bacillus, Azospirillum brasilense, Rhizobium, Enterobacter, Xantomonas, Alcaligenes piechaudii, Agrobacterium Rhizobium spp., leguminosarum, and Comamonas acidovorans spp. [27, PGPRs, which possess enzyme Aminocyclopropane-1-carboxylate (ACC) deaminase, facilitate plant growth and development by lowering ethylene levels, inducing salt tolerance and reducing drought stress in plants [46, 7].

# 3. PGPR as biological control agents

## 3.1. Biological control

Biological control, also known as "biocontrol", is the deliberate use of the biological capabilities (natural mechanisms of action and/or interactions) of a beneficial species to reduce the development of another harmful species [62]. The need for natural, ecologically sustainable, environment-friendly and non-toxic alternatives to chemicals is increasingly being sought, and is leading to the consideration of using PGPRs as biocontrol agents because of the strength that they give to the rhizosphere to deal with threats that target plant roots, in addition to contributing to restoring biodiversity in agro-ecosystems [2].

## 3.2. Potential of PGPRs in biocontrol

Biocontrol can be as effective in controlling plant pathogenic diseases (Table 01) as the use of chemical fungicides [17]. For example, the *Pseudomonas fluorescens* strain can reduce red pepper plant crown rot disease caused by the fungus *Sclerotium rolfsii* by 78% [56]. Also, *Gliocladum vireus* and *Burkholderia cepacia* 

have been used to reduce tomato diseases caused by Sclerotium rolfsii, Pythium ultimum, Rhizoctonia solani and Fusarium oxysporum f. sp. Lycopersici [41].

The particularity of PGPRs, by having a wide range of modes of action and their ability to adapt to the rhizosphere, contribute to biocontrol becoming more sustainable than chemicals. Thus, the use of PGPRs is considered an alternative route to the use of chemicals [12], [8]. In addition, several biocontrol products (Table 02) are marketed and used worldwide [20, 18].

Table 1. Example of biocontrol microorganisms used against plant pathogens [18].

Biocontrol agent	Target Plant Pathogens	Mechanism(s) of action
Bacillus subtilis	Fusarium spp. Erwinia carotovora ssp. Atroseptica and Erwinia	Antibiosis
Bacillus licheniformis P40	carotovora ssp.carotovora	Hydrolytic enzymes
		Competition
Pseudomonas spp. DF-41 and PA-2	Sclerotinia Sclerotiorum	Antibiosis
Streptomyces sp. Di-944	Rhizoctonia solani	Antibiosis
Streptomyces sp. 93	Pythium, Aphanomyces, Phytophtora, Rhizoctonia and Fusarium spp.	Antibiosis
Pseudomonas aeruginosa	Aspergillus flavus, Aspergillus niger, Rhizoctonia bataticola, Rhizoctonia	Antibiosis
	solani, Sclerotium rolfsii and Puccinia arachidis	Competition
Streptomyces diastatochromogenes	Streptomyces scabies	Antibiosis
PonSSII		Competition
Pseudomonas spp.	Gaeumannomyces graminis var. tritici, Pseudomonas tolaasii, Fusarium	Antibiosis
	oxysporum f. sp. Lini and Erwinia amylovora	Competition
Pseudomonas fluorescens	Pectobacterium carotovorum subsp, atrosepticum and Pectobacterium	Hydrolytic enzymes
Pseudomonas putida	carotovorum subsp.carotovorum	Competition Antibiosis
		Induced Resistance System
Trichoderma spp.	Several phytopathogenic fungi	Antibiosis
		Competition
Serratia plymuthica A30	Dickeya solani	Antibiosis

Table 1. Some antagonistic agents marketed as biological products for the treatment of certain plant pathogens [54, 26].

Product	Microorganisms	Diseases treated	Distributor
QA 10	Ampelomyces quisqualis	Mildious	Ecogen, USA
Binab T	Trichoderma spp.	Root rot, fusariosis	Bio innovation AB, New Zealand, USA
Biofox C Fusaclean	Fusarium oxysporum (non-pathogenic)	Fusariosis	S.I.A.P.A., USA
Bio-fungus	Trichoderma harzianum	Root rot, fusariosis	De Ceuster, USA, EU, New Zealand
Intercept	Pseudomonas cepacian	Root rot	Soil Technologies, USA
PSSOL	Ralstonia solanacearum	Root rot	Natural Plant Protection, France
Contans KONI	Coniothyrium minitans	Root rot	Prophyta Biologischer, Hungary, Germany
Polyversum	Pythium oligandrum	Root rot	Biopreparaty, Czech Republic
Primastop (Prestop Mix)	Gliocladium catenulatum	Root rot, fusariosis	Kemira Agro, Finland
Root Shield, Plant Shield, T-22 Planting	Trichoderma harzianum-T22	Root rot	Bioworks, USA, EU, New Zealand
Soil Gard	Gliocladium virens GL-21	Root rot	Therma Trrilogy, USA
Sporodex	Pseudozyma flocculoza	Mildew	Plant products, Canada
Trieco	Trichoderma viride	Root rot, fusariosis	Ecosens Laboratories, India
GBO3, MbI 600	Bacillus subtilis	Melting of seedlings	Horiculture, USA
Mycostop	Streptomyces griseovoridis	Fusariosis, melting of seedlings	Kemira Agro Oy, Helsinki, Finland

# 3.2. Modes of action of PGPRs in biocontrol

The modes of action of PGPRs as a biocontrol agent depend mainly on the microorganism used and the type of plant pathogen to which is applied [11]. In general, the main modes of biocontrol attributed to PGPRs to reduce soil-borne diseases are as follows:

#### 3.2.1. Antagonism

Beneficial rhizobacteria that can secrete substances that inhibit the growth of phytopathogenic microorganisms are called antagonistic bacteria. Therefore, antagonism is the ability of one germ to inhibit the growth of another germ when they are in the same micro-biotope [7]. Similarly, it is expressed in the laboratory when they are grown together in the same Petri dish [7], and it often linked to the phenomenon of antibiosis [11]. Antibiotic production is one of the mechanisms used by PGPRs in the prevention of phytopathogenic attacks and in the suppression of biotic diseases [26]. Regarding the use of PGPRs as a biocontrol tool, both genera *Paenibacillus spp* and *Bacillus* are frequently documented [36].

## 3.2.2. Competition

Competition for space, nutrients or other environmental factors that become limiting to microbial growth is a biological mechanism used by PGPRs to repel or eliminate plant pathogens [55]. An effective competitive agent must be an intense colonizer capable of immediately and efficiently exploiting nutrients present at low concentrations in the soil or stopping their uptaken by other microorganisms [29]. For example, some strains can synthesize extracellular enzymes that led to use organic compounds as a source of energy and/or to degrade phytotoxins [43].

However, in some cases, a reduction in disease may be associated with significant root colonization by PGPRs, which reduces the number of habitable sites for plant pathogenic microorganisms and consequently their growth [47]. The density and intensity of rhizobacteria activity influenced this interaction between beneficial bacteria and phytopathogens [50].

#### 3.2.3. Biofilm formation

Biofilms are structurally complex aggregates of microbial cells attached to a surface and surrounded by an extracellular polymer matrix [57]. PGPRs have a very strong capacity to attach to the plant root system when they form a biofilm [22]. Biofilms have the power to provide significant protection against external aggression and stress, as they act as a protective barrier that prevents the penetration of plant pathogens, releasing a wide range of enzymes, and reducing microbial competition [28]. The best studied examples of PGPRs that form biofilms with Table 03. plants are presented in

Table. 3. Examples of PGPRs involved in biocontrol through biofilm formation.

Bacteria	acteria Features	
Azorhizobium caulinodans	Rice root colonization.	[60]
Azorhizobium brasilense	Root colonization of wheat.	[31]
Acinetobacter calcoaceticus P23	Root colonization of duckweed.	[65]
Bacillus amyloliquefaciens S499	Root colonization of tomato, corn and Arabidopsis thaliana.	[19]
Bacillus polymyxa	Cucumber root colonization.	[66]
Cyanobacteria spp	Improvement of biofilm formation of mixed species with Rhizobium, Azotobacter and Pseudomonas spp.	[51]
Klebsiella pneumoniae	Root colonization of wheat.	[23]
Pantoea agglomerans	Root colonization of chickpeas and wheat.	[10, 4]
Rhizobium leguminosarum bv. viciae 3841	Root colonization of various legumes.	[61, 64]
Rhizobium (Sinorhizobium) sp. strain NGR234	Root colonization of various legumes and competitive colonization in the cowpea rhizosphere.	[37]

#### 3.2.4. Hydrolytic enzymes

The synthesis of hydrolytic enzymes is one of the essential biocontrol mechanisms used by PGPRs against telluric plant pathogens [52]. These strains play a major role in decomposing organic matter in ecosystems and thus protecting plants from environmental stresses [45]. PGPRs can produce certain enzymes, such as amylase [53], chitinase [21], phosphatase [15], protease [16], urease [68], cellulase and lipase [52].

### 3.2.5. Improvement of stress resistance

The action of PGPRs can improve plant's resistance against pathogens. It is mainly due to two signalling pathways (Figure 02):

√The Acquired Systemic Resistance (ASR) whose signal molecule is salicylic acid. It acts by increasing the production of salicylic acid during a microbial infection at the site of contamination as well as in the whole plant. In some plant/pathogen models, salicylic acid, brought exogenously by fluorescent *Pseudomonas*, conferred protection against pathogens [42].

✓Induced Systemic Resistance (ISR): Some PGPRs can stimulate the induced response mechanisms in the plant and lead the whole plant to a state of resistance called Induced Systemic Resistance (ISR) [49, 59].

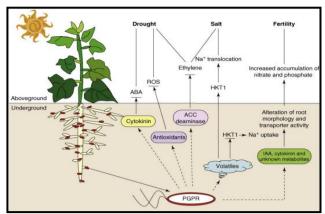


Fig. 2. Roles of PGPRs in promoting plant growth under stress conditions (ABA: abscisic acid; ACC: 1-aminocyclopropane-1-carboxylate; HKT1: high-affinity K+transporter 1; IAA: indole acetic acid; IST: induced systemic tolerance; PGPR: plant growth promoting rhizobacteria; ROS: reactive oxygen species) [67].

### 4. Conclusion

The potential of PGPRs in biocontrol is well established, and their use is proving to be a promising strategy for chemical pesticides. In the present review, the phytoprotective effects of certain PGPRs suggest the possibility of the direct inclusion of these microorganisms in programs for the prevention and control of microbial infections of plants, particularly in agriculture.

#### **Conflict of Interest**

The authors declare that they have no conflict of interest

#### References

- 1. Alderman S.C., Coats D.D. and Crowe F.J. Impact of ergot on Kentucky bluegrass grown for seed in northeastern Oregon. *Plant Dis.* 1996, 80, 853-855.
- 2. Altieri M.A. The ecological role of biodiversity in agroecosystems. Agri. Ecosystems Environ. 1999, 74, 19-31.
- 3. Amellal N., Burtin G., Bartoli F., and Heulin T. Colonization of wheat roots by an exopolysaccharide-producing *Pantoea agglomerans* strain and its effect on rhizosphere soil aggregation. *Appl Environ Microbiol.* 1998, 64, 3740–3747.
- 4. Aouar L. Isolement et identification des actinomycètes antagonistes des microorganismes phytopathogénes. Diplôme de Doctorat en Sciences En Biochimie Microbiologie Appliquées. Université M'entourai-Constantine, Algérie: 2012, p35.
- 5. Benaissa A. Aspects physiologiques et rhizosphériques de *Rhus tripartita (Ucria) Grande* en relation avec l'aridité. « Physiological and rhizospheric aspects of *Rhus tripartita (Ucria) Grande* in relation to aridity". Doctoral dissertation, university of science and technology of Houari Boumediene, Algiers, Algeria; 2020, p65.
- 6. Benaissa A. Plant Growth Promoting Rhizobacteria. A Review, Algerian J. Env. Sc. Technology. 2019, 5(1), 873-880
- 7. Benaissa, A., Djebbar, R and Abderrahmani, A. Antagonistic effect of plant growth promoting rhizobacteria associated with *Rhus tripartitus* on Gram positive and negative bacteria. Analele Universității din Oradea, Fascicula Biologie *Original Paper* Tom. XXVI. 2019, (2), 67-72.
- 8. Benbrook C.M., Groth E., Halloran J.M., Hansen M.K. and Marquardt S. Pest management at the crossroads. Consumers union's of United States Inc. Yonkers. USA; 1996, p272.
- 9. Campell R. and Greaves M.P. Anatomy and community structure of the rhizosphere. In *the rhizosphere*, eds. J. M. Lynch. Wiley Series in Ecological and Applied Microbiologiy, UK; 1990, p11-34
- 10. Chauhan P.S and Nautiyal C.S. The *purB* gene controls rhizosphere colonization by *Pantoea agglomerans*, *Letters in Applied Microbiology*. 2010, 50, 205–210.
- 11. Cherif H. Amélioration de la croissance du blé dur en milieu salin par inoculation avec *Bacillus* sp. et *Pantoea agglomerans* isolées de sols arides. Thèse de doctorat en sciences. Université Ferhat Abbas Sétif 1, Algérie ; 2014, p177.
- 12. Cook R.J. Making greater use of introduced microorganisms for biological control of plant pathogens. *Annu. Rev. Phytopathol.* 1993, 31, 53–80.
- 13. Curl E.A and B., Truelove. The rhizosphere. Journal of nutrition and soil science. Springer-Verlag, Berlin-Heidelberg-New York. 1986, p288.
- 14. De Kouassi M. Les possibilités de la lutte microbiologique. , *VertigO la revue électronique en sciences de l'environnement* [En ligne]. 2001, 2(2). DOI : https://doi.org/10.4000/vertigo.4091
- 15. De T. K., Sarkar T. K., De M., Maity T.K. Mukherjee A., Das S. Abundance and occurrence of phosphate solubilizing bacteria and phosphatase in Sediment of Hooghly estuary, north east coast of Bay of Bengal, India. *Journal of Coastal Development*. 2011, 15(1), 9-16.
- 16. Elad Y and Kapat A. The role of *Trichoderma harzianum* protease in the biocontrol of Botrytis cinerea. European Journal of Plant Pathology. 1999. 105(2), 177-189.
- 17. Emmert E.A.B. and Handelsman J. Biocontrol of plant disease: a (Gram-) positive perspective. *FEMS. Microbiol. Lett.* 1999, 171, 1-9.
- 18. Errakhi R. Contribution d'actinomycètes (Actinobactéries) à la lutte biologique contre *Sclerotium rolfsii* et rôle de l'acide oxalique dans l'induction des mécanismes de défense. Thèse de Doctorat. Université Cadi Ayyad, Marrakech Maroc ; 2008.
- 19. Fan B., Chen X. H., Budiharjo A., Bleiss W., Vater J. and Borriss R. Efficient colonization plant roots by the plant growth promoting bacterium *Bacillus amyloliquefaciens* FZB42, engineered to express green florescent protein, *J of Biotechnol*. 2011,

- 151(4), 303–311. https://doi.org/10.1016/j.jbiotec.2010.12.022
- 20. Fravel D.R. Commercialization and implementation of biocontrol. *Ann. Rev. Phytopathol.* 2005, 43, 337-359. https://doi.org/10.1146/annurev.phyto.43.032904.092924
- 21. Goswami D., Thakker J.N., Dhandhukia P.C. Portraying mechanics of plant growth promoting rhizobacteria (PGPR): A review. Cogent Food Agric. 2016, 2(1). https://doi.org/10.1080/23311932.2015.1127500
- 22. Gupta, G., Snehi, S. K., Singh, V. Role of PGPR in biofilm formations and its importance in plant health. *Biofilms in Plant and Soil Health*. 2017, p27-42
- 23. Iniguez A.L., Dong Y and Triplett E.W. Nitrogen fixation in wheat provided by *Klebsiella pneumoniae* 342, *Mol Plant-Microbe Interact*, 2004, 17(10), 1078–1085. https://doi.org/10.1094/MPMI.2004.17.10.1078
- 24. Ishii H. Impact of fungicide resistance in plant pathogens on crop disease control and agricultural environment. Japan Agricultural Research Quarterly. 2006, 40(3), 205-211. https://doi.org/10.6090/jarq.40.205
- 25. Jan A.T., Azam M., Ali A., Rizwanul-Haq Q.M. Novel approaches of beneficial Pseudomonas in mitigation of plant diseases an appraisal. J Plant Int. 2011, 6(4), 195-205. https://doi.org/10.1080/17429145.2010.541944
- 26. Jijakly M.H. La lutte biologique en phytopathologie, *In*: Phytopathology. Lepoivre. Bruxelles, Belgique. 2003, http://hdl.handle.net/2268/37379
- 27. Karnwal, A., Production of indole acetic acid by fluorescent Pseudomonas in the presence of L-Tryptophan and rice root exudates. J. Plant Pathol. 2009. 91(1), 61-63. https://www.jstor.org/stable/41998574
- 28. Kasim W.A., Gaafar R.M., Abou-Ali R.M., Omar, M.N., Hewait H.M. Effect of biofilm forming plant growth promoting rhizobacteria on salinity tolerance in barley. Ann. Agric. Sci. 2016. 61(2), 217–227. https://doi.org/10.1016/j.aoas.2016.07.003
- 29. Kempf, H. J and Wolf, G. Erwinia herbicola as a biocontrol agent of Fusarium culmorum and Puccinia recondita f. sp. tritici on Wheat. Phytopathology. 1989, 79(2), 990-994.
- 30. Khakipour N., Khvazi K., Mojallali H, Pazira E., Asadirahmani H. Production of auxin hormonr by *Fluorescent pseudomonas*. American-Eurasian Journal Agriculture end Environment Science 2008, 4(6), 687-692
- 31. Kim C., Kecskes M.L., Deaker R.J., Gilchrist K., New P.B and Kennedy I.R. Wheat root colonization and nitrogenase activity by *Azospirillum* isolates from crop plants in Korea, *Can J Microbiol*. 2005, 51(11), 948–956. https://doi.org/10.1139/w05-052
- 32. Kim J., Rees D.C. Nitrogenase and biological nitrogen fixation. *Biochemistry*. 1994, 33(2), 389-97 https://doi.org/10.1021/bi00168a001
- 33. Kloepper J W. Plant-growth-promoting rhizobacteria as biological control agents, In: Soil Microbial Ecology, F.B. Jr., Metting .Marcel Dekker inc., N.Y; 1993, p255-273.
- 34. Kloepper, J. W. and Schroth M. N. Plant growth promoting rhizobacteria on radishes. In Proceedings of the 4th International Conference on Plant Pathogenic Bacteriaed. Station and Pathologic Vegetal and Phytobacteriologic. 1978, 2, 879-882.
- 35. Kloepper J. W., R. Rodriguez-Ubana G. W. Zehnder J. F. Murphy E. Sikora, and C. Fernandez. Plant root-bacterial interactions in biological control of soilborne diseases and potential extension to systemic and foliar diseases. Austral. Plant Pathol. 1999, 28, 21–26. https://doi.org/10.1071/AP99003
- 36. Kokalis-Burell N., Kloepper J.W., Reddy MS. Plant growth-promoting rhizobacteria as transplant amendments and their effects on indigenous rhizosphere microorganisms. *Appl Soil Eco.* 2005, 31(1-2), 91-100. https://doi.org/10.1016/j.apsoil.2005.03.007
- 37. Krysciak D., Schmeisser C., Preuss S., Riethausen J., Quitschau M., Grond S. Involvement of multiple loci in quorum quenching of autoinducer I molecules in the nitrogen-fixing symbiont *Rhizobium* (*Sinorhizobium*) sp. strain NGR234, *Appl Environ Microbiol*. 2011, 77(15), 5089–5099. http://dx.doi.org/10.1128/AEM.00112-11.
- 38. Leong J. Siderophores: their biochemistry and possible role in the biocontrol of plant pathogens. Annu Rev Phytopathol. 1986, 24, 187-208. https://doi.org/10.1146/annurev.py.24.090186.001155
- 39. Lepoivre P. Phytopathologie: bases moléculaires et biologiques des pathosystèmes etfondements des stratégies de lutte. De BoeckUniversité, Bruxelles, Belgium. 2003, p432.
- 40. Loper, J. E and Schroth, M. N. Influence of bacterial sources of indole-3-acetic acid on root elongation of sugar beet. *Phytopathology.* 1986, 76 (4),386-389.
- 41. Mao W., Lewis A., Lumsden R.D. and Hebbar K.P. Biocontrol of selected soilborne diseases of tomato and pepper plants. *Crop Prot.* 1998, 17(6), 535-542. https://doi.org/10.1016/S0261-2194(98)00055-6
- 42. Maurhofer M., Reimmann C., Schmidli-Sacherer P., Heeb S., Haas D and Defago G. Salicylic acid biosynthetic genes expressed in *Pseudomonas fluorescens* strain P3 improve the induction of systemic resistance in tobacco against tobacco necrosis virus. *Phytopathology*. 1998, 88(7), 678-684. https://doi.org/10.1094/PHYTO.1998.88.7.678
- 43. Mccarthy A.J., Williams, S.T. *Actinomycetes* as agents of biodegradation in the environment. Gene. 1992, 115(1-2), 189-192. https://doi.org/10.1016/0378-1119(92)90558-7
- 44. MOUAS BOURBIA, Sophia. *Biodisponibilité du potassium dans la rhizosphère d'Olea europaea L.* Thèse de doctorat. Thèse doctorat-Université Mouloud Mammeri-Tizi Ouzou. Algérie ; 2013, p19-20.
- 45. Nadeem S.M., Naveed M., Zahir Z.A., Asghar H.N. Plant–Microbe Interactions for Sustainable Agriculture: Fundamentals and Recent Advances. In: Arora N. (eds) Plant Microbe Symbiosis: Fundamentals and Advances. Springer, New Delhi; 2013, p51-103. https://doi.org/10.1007/978-81-322-1287-4\_2
- 46. Nadeem, S.M., Zahir, Z.A., Naveed, M and Arshad, M. Preliminary investigations on inducing salt tolerance in maize through inoculation with rhizobacteria containing ACC deaminase activity. Can. J. Microbiol. 2007. 53(10): 1141–1149. https://doi.org/10.1139/W07-081
- 47. Nguyen-the C., Carlin F. The microbiology of minimally processed fresh fruits and vegetables. Crit. Rev. Food Sci. Nutr. 1994, 34(4), 371–401. https://doi.org/10.1080/10408399409527668
- 48. Piano S., Neyrotti V., Migheli Q., Gullino M.L. Biocontrol capability of Metschnikowia pulcherrima against Botrytis postharvest rot of apple. *Postharvest Biol. Technol.* 1997, 11 (3), 131-140. https://doi.org/10.1016/S0925-5214(97)00022-7
- 49. Pieterse C. M. J., Van Wees S.C. M., Hoffland E., Van Pelt J.A., Van Loon L. C. Systemic resistance in Arabidopsis induced by biocontrole bacteria is independent of salycilic acid accumulation and pathogenesis- related gene expression. The Plant Cell.

- 1996, 8, 1225-1237. DOI: https://doi.org/10.1105/tpc.8.8.1225
- 50. Podile AR and Kishore KG. Plant growth promoting rhizobacteria. In: Gnanamanickam SS (Ed.). Plant-Associated Bacteria, Springer, Dordrecht; 2006, p195–230.
- 51. Prasanna R., Pattnaik S., Sugitha T.C., Nain L and Saxena A.K. Development of cyanobacterium-based biofilms and their in vitro evaluation for agriculturally useful traits, *Folia Microbiol (Praha)*. 2011, 56, 49–58. https://doi.org/10.1007/s12223-011-0013-5
- 52. Raafat D., Sahl H. G. Chitosan and its antimicrobial potential a critical literature survey. *Microb. Biotechnol.* 2009, 2(2), 186–201. https://doi.org/10.1111/j.1751-7915.2008.00080.x
- 53. Raj S. V., Raja A.K., Vimalanathan A.B., Tyagi M.G., Shah N.H., Johnson Amala Justin N.A., Santhose B.I., Sathiyaseelan K. Study of starch degrading bacteria from kitchen waste soil in the production of Amylase by using paddy straw. *Recent Research in Science and Technology*, 2009, 1(1), 008–013.
- 54. Sabaratnam S. and Traquair J.A. Formulation of a *Streptomyces* biocontrol agent for the suppression of *Rhizoctonia* damping-off in tomato transplants. *Biol. Control.* 2002, 23(3), 245-253. https://doi.org/10.1006/bcon.2001.1014
- 55. Shameer S., Prasad T.N.V.K.V. Plant growth promoting rhizobacteria for sustainable agricultural practices with special reference to biotic and abiotic stresses. *Plant Growth Regul*. 2018, 84, 603–615. https://doi.org/10.1007/s10725-017-0365-1
- 56. Singh A., Mehta S., Singh H.B. and Nautiyal C.S. Biocontrol of collar rot disease of betelvine (*Piper betle L.*) caused by *Sclerotium rolfsii* by using rhizosphere-competent *Pseudomonas fluorescens* NBRI-N6 and *P. fluorescens* NBRI-N. *Cur. Microbiol.* 2003, 47, 153-158. https://doi.org/10.1007/s00284-002-3938-8
- 57. Souza, R. de, Ambrosini, A., and Passaglia, L.M.P. Plant growth-promoting bacteria as inoculants in agricultural soils. *Genet. Mol. Biol.* 2015. 38(4): 401–419. https://doi.org/10.1590/S1415-475738420150053
- 58. Thakore Y., The biopesticide market for global agricultural use, industrial biotechnology. 2006, 2(3), 294-208. https://doi.org/10.1089/ind.2006.2.194
- 59. Van Loon, L. C., Bakker, P and Pieterse, C. M. J. Systemic resistance induced by rhizosphere bacteria. Annual Review of Phytopathology. 1998, 36, 453-483. https://doi.org/10.1146/annurev.phyto.36.1.453
- 60. Van Nieuwenhove C., Holm V., Kulasooriya S.A and Vlassak K. Establishment of *Azorhizobium caulinodans* in the rhizosphere of wetland rice (*Oryza sativa* L.), *Biol Fertil Soils*. 2000, 31, 143–149. https://doi.org/10.1007/s003740050637
- 61. Vanderlinde E.M., Muszynski A., Harrison J.J., Koval S.F., Foreman D.L and Ceri H., *Rhizobium leguminosarum* biovar viciae 3841, deficient in 27-hydroxyoctacosanoatemodified lipopolysaccharide, is impaired in desiccation tolerance, biofilm formation and motility, *Microbiol*. 2009. 155, 3055–3069. doi: 10.1099/mic.0.025031-0
- 62. Vinodkumar S, Nakkeeran S, Renukadevi P and Mohankumar S. Diversity and antiviral potential of rhizospheric and endophytic *Bacillus* species and phyto-antiviral principles against tobacco streak virus in cotton. Agriculture, Ecosystems & Environment. 2018, 267, 42-51. https://doi.org/10.1016/j.agee.2018.08.008
- 63. Weyens N., S. Monchy, S., Vangronsveld, J., Taghavi, S and Vander Lelie., D. PlantMicrobe Partnerships. In K.N. Timmis (ed.), Handbook of hydrocarbon and lipid microbiology, Springer-Verlag, Berlin Heidelberg; 2010, p254-256.
- 64. Williams A., Wilkinson M., Krehenbrink D.M., Russo A., Zorreguieta and J.A Downie, Glucomannan-mediated attachment of *Rhizobium leguminosarum* to pea root hairs is required for competitive nodule infection, *J Bacteriol*. 2008, 190(13), 4706–4715. DOI: 10.1128/JB.01694-07
- 65. Yamaga F., Washio K., Morikawa M. Sustainable biodegradation of phenol by *Acinetobacter calcoaceticus* P23 isolated from the rhizosphere of duckweed *Lemna aoukikusa*, *Environ Sci Technol*. 2010, 44 (16), 6470–6474. https://doi.org/10.1021/es1007017
- 66. Yang J., Kharbanda P.D and Mirza M. Evaluation of *Paenibacillus polymyxa* PKB1 for biocontrol of Pythium disease of cucumber in a hydroponic system, *Acta Hortic (ISHS)*. 2004, 635, 59–6.
- 67. Yang J., Kloepper J.W and Ryu C.M. Rhizosphere bacteria help plants tolerate abiotic stress. *Trends in Plant Science*. 2009, 14 (1), 1-4. https://doi.org/10.1016/j.tplants.2008.10.004
- 68. Yang Z, Liu S, Zheng D and Feng S. Effects of cadium, zinc and lead on soil enzyme activities. J. Environ. 2006, 18 (6), 1135-1141. https://doi.org/10.1016/S1001-0742(06)60051-X
- 69. Zahir, Z.A., Munir, A., Asghar, H.N., Shaharoona, B and Arshad, M. Effectiveness of rhizobacteria containing ACC-deaminase for growth promotion of pea (Pisum sativum) under drought conditions. J. Microbiol. Biotechnol. 2008, 18(5), 958–963.

### **Recommended Citation**

Bestami M., Ben Malek R., Fellan K and Benaissa A. Biological control by Plant Growth Promoting Rhizobacteria. A review. Algerian Journal of Biosciences. 2020, 01;02:030-036.

DOI: http://dx.doi.org/10.5281/zenodo.4393567



This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License