

Physicochemical Characterization and Antimicrobial Activity of Essential Oil from *Rosmarinus officinalis* L, Growing in the Wild

West Saharan Atlas of Algeria

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Abstract - The antimicrobial activity of *Rosmarinus officinalis* L essential oil has been recognized for a long time. In the present study, the antimicrobial properties of the essential oils obtained from the leaves of *R. officinalis* L, have been investigated against six strains of bacteria and six fungi strains.

R. officinalis L, provided a yield of 1.5% of essential oil. The Physicochemical characterization shows that refractive index is about (1.467). Specific density at 20 °C (0.949), optical rotation (+11.7), colour and odour were also determined.

R. Officinalis L. essential oil exhibited stronger antimicrobial activity, with a maximum inhibition zone of 28 mm against *E. Faecalis*. In addition, the essential oil has a remarkable antifungal activity. *P. Escpansum* is the most sensitive strain with an inhibition diameter of 24 mm. All fungal strains were inhibited at a low concentration of 1/600.

Keywords: Essential oil, *Rosmarinus officinalis* L, antimicrobial activity, physicochemical characterization.

1.Introduction

Essential oils and extracts from aromatic plants have long been used for a wide variety of medicinal and domestic purposes. Antimicrobial properties of essential oils obtained from aerial parts and

seeds of some aromatic plants such as *Matricaria pubescens* (Desf.), *Rosmarinus officinalis* L and *Artemisia herba alba* have been investigated and reviewed against food-related microorganisms as well as their applications in food systems (Gachkar et al.,2007 ; Atik bekkara et al.,2007;

Chanthaphon *et al.*, 2008 ; Makhloufi *et al.*, 2012 ; Makhloufi *et al.*, 2015).

Rosemary has been the subject of recent research in the pharmaceutical and food industries. It has excellent antioxidant and antimicrobial properties, due to certain compounds (carnosol, carnosic acid, ursolic acid, betulinic acid, and the rosmaridiphenol rosmanol) (Thoresen *et al.*, 2003; Boutabia *et al.*, 2016).

R. officinalis L. is a shrub belonging to the Lamiaceae family, in the Mediterranean area, including Algeria, which lies on the arid slopes and hills (Dellile, 2007; Makhloufi, 2012). In the west Saharan Atlas of Algeria, it is quite frequent in the mountains of Mourghad, Mekthern, Bouaboud, and Founassa, etc.

2. Materials and Methods

R. officinalis L. specimens were collected from Morghad Mountain in the region of Founassa (32°30'43.047" N de latitude, 0°50'9.669" O), Saharan Atlas, West of Ain Sefra (Figure 1) during December 2016, and January 2017. These biomasses were dried for fifteen days in the dark at an ambient laboratory temperature (20-28 °C).

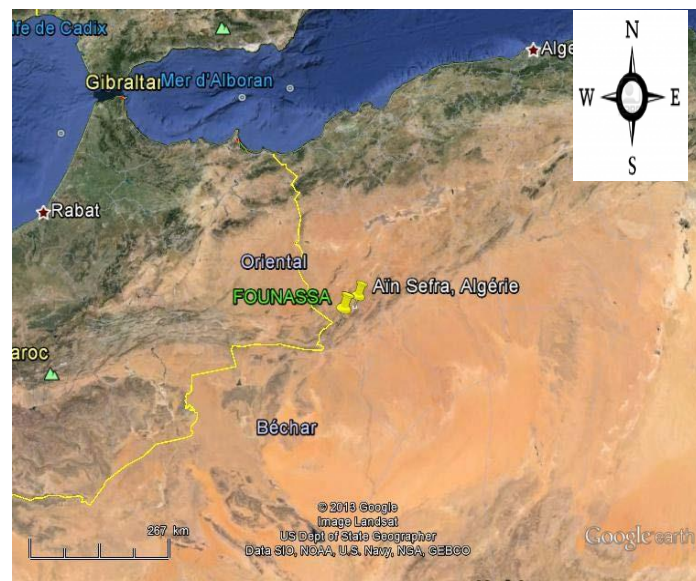


Figure 1: Geographical Situation of the Study Area (Founassa) (Bouyahiaoui, 2017)

2.1. Distillation of the Essential Oil

The dried aerial parts were ground before the operation, and then, 100 g of ground *R. officinalis* L were submitted to hydrodistillation for 3h using a Clevenger apparatus (Amarti *et al.*, 2008; Makhloufi *et al.*, 2011; Akermi *et al.*, 2017). The distilled essential oils were dried over anhydrous sodium sulphate, filtered and stored at +4 °C until it was used (Chanthaphon *et al.*, 2008; Ayoughi *et al.*, 2011).

2.2. Physical and Chemical characterization of essential oil *R. officinalis* L.

Methods conform to A.F.N.O.R.(1986) have determined the specific density at 20°C ; Optical Rotation (OR) was measured with a polarimeter. The refractive index (RI) was measured with a refractometer at

20°C. The methods used for the determination of the acid index and ester are also used following the AFNOR standards.

2.3 Antimicrobial activity

2.3.1 Microbial strains

The antimicrobial activity was evaluated by paper disc diffusion and dilution methods against six selected fungi, namely: *Aspergillus niger*, *A.flavus*(1) , *Penicillium purpurogenum*(Isolated from dates), *P.Jensinii*, *P.escpansum* and , *A. flavus* (2) isolated from Wheat, and six selected Gram-positive and Gram-negative species: *Escherichia coli* ATCC 25922, *Staphylococcus aureus* ATCC 25923, *Pseudomonas aeruginosa* ATCC27853, *Bacillus cereus* ATCC 11778, *Enterococcus feacalis* ATCC 29212 and *Listeria monocytogenes* ATCC19115.

2.3.2 Disc diffusion method

The agar disc diffusion method was employed for the determination of antifungal activities of the essential oil in question. Briefly, the fungal cultures were grown on PDA. The 7-day old culture of mycelial mat was washed, and then suspended in normal saline solution. The colony forming units (CFU/ml) of suspension of the test fungus was determined and test inoculum was adjusted to 10^6 CFU/ml. These conidia were used for antifungal essay tests. Inocula (0.1ml) were

applied on the surface of the PDA plate and spread using a sterile glass spreader (Bansod & Rai, 2008). The qualitative antibacterial was performed using culture growth at 37 °C for 18 h and adjusted to approximately 10^8 colony forming unit per milliliter (CFU/ml). The culture medium used for the bacteria was Mueller Hinton Agar (MHA) (Gachkar et al., 2007). Five hundred microliters of the inoculums were spread over plates containing MHA and a Whatman paper disc (6 mm in diameter) were impregnated with 10 µl of the undiluted oil and were placed on the inoculated plates. The plates were left for 30 min at room temperature, and incubated at 37 °C for 24 h (Shunying et al., 2005; Bekhchi et al., 2008; bourkhiss et al., 2007). The diameters of the inhibition zones were measured in millimeters. Control assay discs were also used; all tests were performed in triplicate (Yesil et al., 2007).

2.3.3 Dilution method (MIC)

Antimicrobial tests were performed according to the method reported by Remmal et al. (1993), and Farah et al. (2001). The essential oil is emulsified with an agar solution of 0.2% in order to disperse the compounds and improve their contact with the tested germs, before being diluted to one tenth in the agar solution. Quantities of this dilution were added to

test tubes containing Mueller Hinton agar for bacteria, and PDA for fungi. The final concentrations of essential oil are from 1 / 100, to 1 / 1000 (v / v). In parallel, Control assay containing only the culture medium and agar solution at 0.2% were also used. The MIC is the lowest concentration of essential oil giving no visible growth to the naked eye (Makhloufi *et al.*, 2011).

3. Results and discussion

The essential oil, with pale yellow colors, was obtained by hydrodistillation from *R.*

officinalis L with the yield of 1.5 % (v/w) on dry weight basis.

This yield is comparable to that quoted by Hilan *et al.* (2006) (1.52%) and is higher than that quoted by Bekkara Atik *et al.* (2007) (0.8%), Biljana *et al.* (2007) (1.18 %). In contrast, this performance was revealed lower than that given by the same species from different regions such as Sardinia whose performance can reach 1.75% (Angioni *et al.*, 2004).

Table 01: Physico-chemical index of essential oil of *R. officinalis* L.

Refractive index	Optical Rotation	Specific density at 20 °C	Ester index	Acid index
1.467	+11.70	0.949	18.50	0,550

As can be seen from Table 1, the major differences were found in density and Refractive index. The main properties of *R. officinalis* of Campestre da Serra (Rio Grande do Sul State) : The physico-chemical parameters averaged 0.8887 g/cm³ for specific gravity, 1.4689 for refractive index, and +11.82° for optical rotation, and there were no significant variations in either the chemical or physico-chemical data in the different years (Atti-Santos, 2005), for Morocco the density is about 0.898 and the Refractive index equals 1.468. (EL Kamli *et al.*, 2017)

The Physico-chemical properties of *R. officinalis* essential oil and yield and chemical composition depend on environmental factors which include cultivation and habitat climate conditions (temperature, humidity, radiation, wind, soil properties, geographical location, and harvest time and methods), and postharvest techniques .

The Physico-Chemical Characteristics Value of British Pharmacopoeia (2008) are: Relative density 0.895 to 0.920, Refractive index 1.464 to 1.473, Optical

rotation -5 to 8 , the Appearance between Colourless to pale yellow liquid .

3.1. Antimicrobial activity

3.1.1. Disc diffusion assay

The growth inhibition zones measured by disc diffusion method are presented in Fig. 2 and 3. According to these results, the essential oil of *R. officinalis L.* have great antimicrobial activity against most of the investigated strains. The diameters of growth inhibition zone ranged from 16 to 28 mm (including the diameter of the disc- 6 mm) with the highest inhibition zone values observed against *E. faecalis*

(28mm)for Bacteria and *P.escpansum* (24mm) for fungi.

As For the direct contact method, the results differ depending on the seeds used (Table 2 and 3). in vitro, *R. officinalis L.* essential oil proved to have a good inhibitory activity against the tested germs. However, the microorganisms studied did not show the same sensitivity against the essential oil. Rosemary essential oil inhibited both fungal agents tested at a concentration of 1/600 v/v.

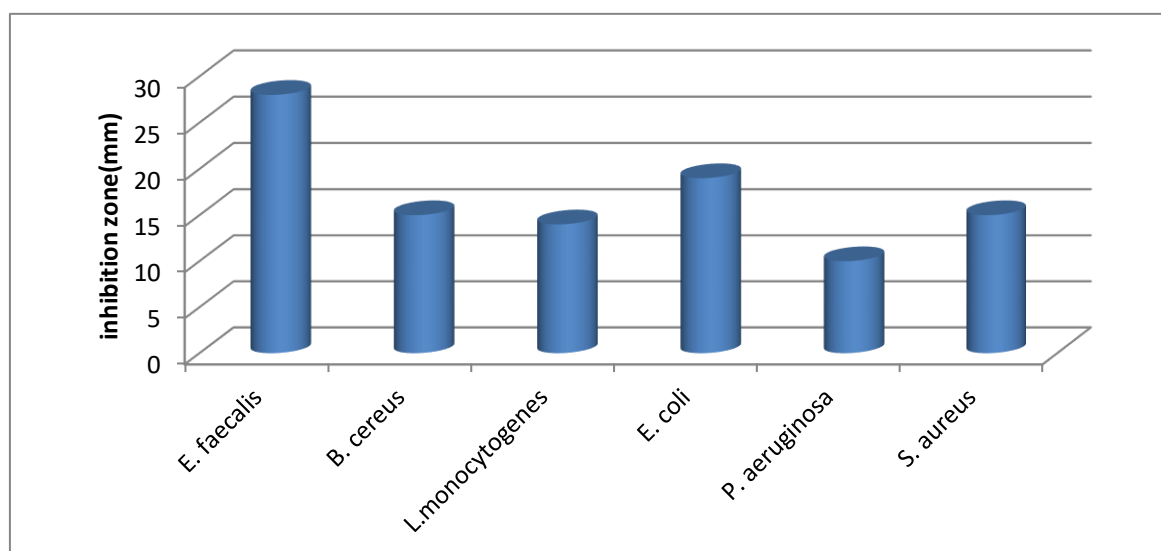


Figure 02 : Antibacterial activity of essential oil (EO) by disc diffusion assay

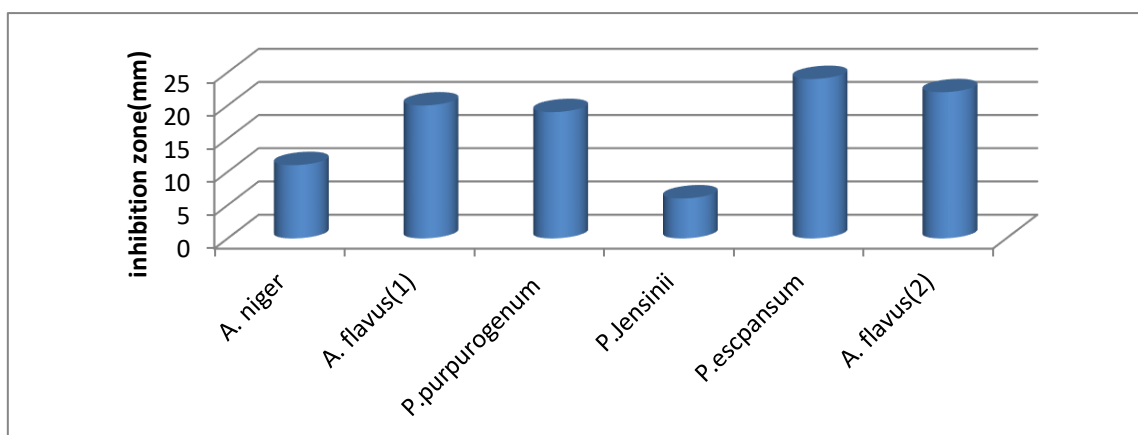


Figure 03: Antifungal activity of essential oil (EO) by disc diffusion assay

Table 02: MIC of essential oil (EO)(v/v) on bacteria strains

Concentrations Bacteria	1/10	1/25	1/50	1/75	1/100	1/125	1/150	1/175
<i>P.aeruginosa</i>	-	-	-	+	+	+	+	+
<i>S.aureus</i>	-	-	+	+	+	+	+	+
<i>B.cereus</i>	-	-	+	+	+	+	+	+
<i>L.monocytogenes</i>	-	-	-	+	+	+	+	+
<i>E.faecalis</i>	-	-	-	-	-	+	+	+
<i>E.coli</i>	-	-	+	+	+	+	+	+

Table 03: MIC of essential oil (EO)(v/v) on fungal strains

Concentrations Fungi	1/500	1/550	1/600	1/750	1/800	1/850	1/900	1/950
<i>A.niger</i>	-	-	-	+	+	+	+	+
<i>A.flavus(1)</i>	-	-	-	+	+	+	+	+
<i>P.purpurogenum</i>	-	-	-	-	+	+	+	+
<i>P.jensinii</i>	-	-	-	-	+	+	+	+
<i>A.flavus(2)</i>	-	-	-	+	+	+	+	+
<i>P.escansum</i>	-	-	-	-	+	+	+	+

(-):Inhibition (+): Growth

The data indicated that Gram-positive bacteria were more sensitive to essential oils than to Gram-negative bacteria due to their outer membrane barriers (Burt, 2004). Among Gram-positive bacteria, *E. faecalis* was the most sensitive to essential oil from rosemary (MIC 1/100 v/v) whereas *S.aureus* was the most resistant (MIC 1/25v/v). Several researchers reported that *R. officinalis* L essential oil inhibited some Gram-positive and negative bacteria (*S.aureus*, *L.monocytogenes*, *E.coli*....) (Mathlouthi *et al.*, 2009; Rozman and Jeršek, 2009). Moreno *et al.* (2006) reported that rosemary plants are rich sources of phenolic compounds with high antimicrobial activity against both Gram-positive and Gram-negative bacteria. High percent of the antimicrobial activity they attributed to carnosic acid and carnosol.

4. Conclusion

In conclusion, Hydro distilled-essential oil from *R. officinalis* L. has a good inhibitory activity. The strong antifungal activity of *R. officinalis* L. against filamentous fungi strains is an indication of the broad spectrum of the oil antifungal potential. This Activity could be a natural alternative to synthetic antimicrobial and preservatives to enhance the safety and the shelf life of food.

5. Acknowledgments

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